

RESEARCH

Treating infant colic with the probiotic *Lactobacillus reuteri*: double blind, placebo controlled randomised trial

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Abstract

Objective To determine whether the probiotic *Lactobacillus reuteri* DSM 17938 reduces crying or fussing in a broad community based sample of breastfed infants and formula fed infants with colic aged less than 3 months.

Design Double blind, placebo controlled randomised trial.

Setting Community based sample (primary and secondary level care centres) in Melbourne, Australia.

Participants 167 breastfed infants or formula fed infants aged less than 3 months meeting Wessel's criteria for crying or fussing: 85 were randomised to receive probiotic and 82 to receive placebo.

Interventions Oral daily *L reuteri* (1×10^8 colony forming units) versus placebo for one month.

Main outcomes measures The primary outcome was daily duration of cry or fuss at 1 month. Secondary outcomes were duration of cry or fuss; number of cry or fuss episodes; sleep duration of infant at 7, 14, and 21 days, and 1 and 6 months; maternal mental health (Edinburgh postnatal depression subscale); family functioning (paediatric quality of life inventory), parent quality adjusted life years (assessment of quality of life) at 1 and 6 months; infant functioning (paediatric quality of life inventory) at 6 months; infant faecal microbiota (microbial diversity, colonisation with *Escherichia coli*), and calprotectin levels at 1 month. In intention to treat analyses the two groups were compared using regression models adjusted for potential confounders.

Results Of 167 infants randomised from August 2011 to August 2012, 127 (76%) were retained to primary outcome; of these, a subset was analysed for faecal microbial diversity, *E coli* colonisation, and calprotectin levels. Adherence was high. Mean daily cry or fuss time fell

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Extra material supplied by the author (see <http://www.bmj.com/content/348/bmj.g2107?tab=related#webextra>)

Total daily infant cry or fuss time through study period and at 6 months

Video on bmj.com (see also <http://bmj.com/video>)



Video abstract

steadily in both groups. At 1 month, the probiotic group cried or fussed 49 minutes more than the placebo group (95% confidence interval 8 to 90 minutes, $P=0.02$); this mainly reflected more fussing, especially for formula fed infants. The groups were similar on all secondary outcomes. No study related adverse events occurred.

Conclusions *L reuteri* DSM 17938 did not benefit a community sample of breastfed infants and formula fed infants with colic. These findings differ from previous smaller trials of selected populations and do not support a general recommendation for the use of probiotics to treat colic in infants.

Trial registration Current Controlled Trials ISRCTN95287767.

Introduction

Infant colic, or excessive crying of unknown cause, affects up to 20% of infants¹ and is a major burden to families and health services. Infant colic is often defined by the Wessel's criteria of crying or fussing for three hours or more a day for three days or more per week for three weeks in infants aged less than 3 months.² Although infant colic spontaneously resolves after the first three to four months after birth, it is associated with maternal depression,³ early breastfeeding cessation,⁴ and shaken baby syndrome.⁵ Infant distress is one of the most common presenting problems to primary, secondary, and tertiary healthcare sectors, costing the UK healthcare system millions of pounds annually.⁶

The cause of infant colic remains elusive despite decades of research. Psychosocial hypotheses include poor maternal-infant interactions, maternal anxiety and depression, and difficult infant temperament.⁷ Gastrointestinal theories include increased intra-abdominal gas, hyperperistalsis, and visceral pain.⁷ One study has suggested that infants with colic may have increased faecal calprotectin levels,⁸ suggesting a possible role for gut inflammation; however, another study suggested no differences in faecal calprotectin levels between infants with and without colic.⁹

No single effective treatment for colic exists, and most clinical guidelines recommend support and reassurance as the mainstay of management.¹⁰⁻¹³ The use of hypoallergenic formulas or elimination of cow's milk protein from the diet of mothers who are breast feeding may possibly be effective,¹⁰⁻¹² yet not all irritable infants respond.¹⁴⁻¹⁵ Anticholinergic drugs (for example, dicycloverine (dicyclomine)) are effective¹⁶⁻¹⁸ but have potentially dangerous side effects, including breathing difficulties and coma.¹⁹ An effective, practical, and acceptable intervention for infant colic would represent a major advance in clinical and public health.

Recently, research into the use of probiotics (live micro-organisms that confer a health benefit) for colic has been rapidly gaining momentum. Infants with colic are reported to have increased concentrations of gas forming organisms and proteobacteria such as *Escherichia coli* in their gut.^{8,22} Colonisation with certain intestinal micro-organisms, such as *Bifidobacterium* and *Lactobacillus* species, along with increased intestinal microbial diversity, may protect against infant distress.^{8,23} Probiotics enhance the mucosal barrier and promote microbial diversity in the gut.²⁴⁻²⁵ They may reduce concentrations of proteobacteria and gas forming coliform and reduce intestinal inflammation.²⁶⁻³⁰ A recent meta-analysis of three small, randomised controlled trials of breastfed infants with colic reported that *Lactobacillus reuteri* noticeably reduced crying time at 21 days post supplementation.³¹ However, one trial was unblinded,³² two included only infants with mothers on dairy-free diets,³²⁻³³ and none used validated measures of infant distress. No trials have included formula fed infants,

which is relevant because colic is associated with early breastfeeding cessation.⁴ Despite these major limitations, the use of probiotics for colic has been rapidly taken up internationally. An urgent need exists for a larger, more rigorous trial that includes infants unselected for feeding method to clarify whether *L reuteri* is effective for infant colic in the general population.

We determined whether the probiotic *L reuteri* DSM 17938 benefited infants aged less than 3 months (<13 weeks) with colic, irrespective of feeding mode. We also examined its effect on gut microbiota, faecal calprotectin levels, and *E coli* colonisation, all implicated in the mechanism of disease. We hypothesised that compared with the placebo (control) group, the *L reuteri* (treatment) group would show lower mean daily cry or fuss time at 1 month (primary outcome), and at 7, 14, and 21 days, and 6 months, and fewer daily episodes of cry or fuss (7, 14, 21 days, 1 and 6 months); longer infant sleep duration (7, 14, 21 days, 1 and 6 months); better mean scores on a standardised measure of maternal mental health; better mean scores on a standardised measure of infant (6 months) and family functioning (1 and 6 months); better mean scores on a standardised measure of parent quality adjusted life years (1 and 6 months); and changes in gut microbiota (increased faecal microbial diversity, less *E coli* colonisation (1 month), and reduced faecal calprotectin levels (1 month)). We hypothesised that these effects would be sustained within the breastfed group and formula fed group of infants.

Methods

Trial design

This was a phase III, double blind, randomised placebo controlled trial. Details of the protocol are described elsewhere.³⁴

Participants

Between August 2011 and August 2012 we recruited participants from a range of services accessed by carers of newborn babies in Melbourne, Australia: the Royal Children's Hospital emergency department, the Royal Children's Hospital Unsettled Babies Clinic (a referral based outpatient clinic), Tweddle Child and Family Health Centre (a mother-infant parenting centre), two maternal child health centres (universal nurse health checks), and paediatricians at the Royal Children's Hospital and in private practices. Families could also contact the study team to be involved.

We recruited healthy term infants less than 13 weeks of age with infant colic, defined by modified Wessel's criteria of crying or fussing for three hours or more a day for three days or more over seven days. The study doctor assessed eligibility through a structured telephone or face to face interview. Fussing was defined as "behaviour that is not quite crying but not awake and content either."³⁵ We excluded infants who weighed less than 2500 g at birth; failed to thrive; had major medical problems; had an allergy to cow's milk protein (confirmed by symptom resolution after a two week trial of hypoallergenic formula or a cow's milk protein-free diet in mothers of breastfed infants); or took solids, antibiotics, or *L reuteri* at the start of the trial. We also excluded breastfed infants whose mothers took *L reuteri* at trial start, and caregivers with insufficient English to complete questionnaires.

Intervention

The treatment was *L reuteri* DSM 17938 (0.2×10⁸ colony forming units per drop) in an oil suspension, given once daily

(five drops orally) for one month. The placebo was maltodextrin in the same oil suspension with the same appearance, colour, and taste as the treatment, identically packaged and stored. Caregivers were instructed to dispense five drops of study liquid onto a spoon to administer to their infants at the same time each day. The dosing of *L reuteri* is the same as that used in the three previous trials showing effectiveness of the probiotic in managing colic.³²⁻³⁶

Randomisation, allocation concealment, blinding

Randomisation was stratified by method of infant feeding (breast fed v formula fed) and age (≤ 6 weeks v > 6 weeks, owing to the natural peak in crying at around six weeks of age). We allocated infants who were both breast fed and formula fed to the formula fed stratum. An independent statistician prepared the computer generated randomisation schedule using a block size of two to maintain balance between treatment arms within each stratum. The trial was double blind, whereby treatment allocation was concealed from all study investigators and participants at every phase, including measurement of outcomes. Study investigators allocated infants to one of four randomisation strata according to their age and feeding type. An independent pharmacist (who was not part of the study team and had no access to the study database) then allocated the study identification number to each infant and dispensed the study product according to the randomisation schedule. This randomisation schedule was not revealed to the study investigators until all outcomes analyses at one month were complete. The treatment and placebo drops were packaged identically such that study investigators and families were blinded to treatment allocation throughout the study.

Outcomes, adherence, adverse events

Table 1¹ shows the outcome measures at the different time points. Funding allowed analysis of a subset of faecal samples for microbial diversity, *E coli* load, and calprotectin levels collected at one month post-intervention. We measured adherence by weekly parental report using a diary, with non-adherence defined as carers reporting discontinuing the intervention within two weeks of trial start. We weighed all bottles before and after intervention to determine equivalence in adherence between the groups. We also assessed for *L reuteri* in 65 faecal samples. Parents recorded any adverse events weekly. For the six month outcomes, we allowed the last 91 carers in the trial to fill in one day of the study diary instead of two to reduce the study burden and improve retention rate.

Statistical analyses

Sample size estimation

A sample size of 160 provided 80% power to detect a minimum effect size of 0.5 standard deviations difference in the mean daily cry or fuss time between treatment groups with a significance level of $P < 0.05$, allowing for a dropout rate of 20% (target sample size of 128 for primary analysis). It also provided 80% power to detect a minimum effect size of 0.85 standard deviations difference in the mean daily cry or fuss time between treatment groups within either the formula fed infants or the breastfed infants, assuming 40% of infants in the sample were exclusively breast fed.

Analyses

We used Stata 12.0 to carry out analyses using the intention to treat principle. If more than 30% of data were missing in the

diary at one month, we excluded the family from analysis. We considered families who lost their diaries as being lost to follow-up because they had no cry or fuss data throughout the whole study period. In unadjusted analyses, we compared all outcomes by *t* test and non-parametric tests for continuous data, and χ^2 tests for dichotomous data. The primary outcome was the daily duration of cry or fuss at one month. In addition, we analysed a dichotomised indicator of treatment response, defined as a 50% reduction in cry or fuss time. We planned subgroup analyses a priori to examine treatment differences separately among breastfed infants and formula fed infants, and among infants with and without a family history of atopy.

We used regression models to estimate treatment effects, as mean differences with 95% confidence intervals, adjusting for potential confounding factors identified a priori and measured at baseline. These were infant age and sex, mode of delivery, type of feeding, family history of atopy, and the respective baseline variable (for example, baseline duration of cry or fuss, number of episodes of cry or fuss, maternal depression). We repeated the regression models using bootstrapping for skewed data. For further longitudinal analyses examining trends in cry or fuss time within individual babies we used random effect regression models.

Results

Participant flow and baseline characteristics

Overall, 167 families were randomised (51% of 329 eligible families; 85 probiotic, 82 placebo). Retention to primary outcome was 76% (127 infants analysed, 67 probiotic, 60 placebo); 11 and 29 families were excluded for loss to follow-up and missing data, respectively (fig 1¹). Other than more boys in the placebo group, the baseline characteristics of both groups were similar (table 2¹). Recruitment occurred mainly from the Royal Children's Hospital emergency department (73%).

Primary outcome

Figure 2¹ plots the steady decline in daily duration of infant cry or fuss in both groups over the study period (see supplementary table 1 for values). Table 3¹ shows and compares all primary and secondary outcomes at 1 and 6 months.

Overall, the two groups were similar on almost all outcomes. At 1 month, the probiotic group cried or fussed an adjusted mean of 49 minutes/day more than the placebo group (95% confidence interval 8 to 90, $P = 0.02$). This was mainly due to more fussing, not more crying, in the probiotic group (adjusted mean difference 52 minutes/day, 19 to 84, $P = 0.002$) at 1 month. The probiotic group fussed significantly more than the placebo group at all time points from day 7 to 1 month (see supplementary table 1). Although daily duration of cry or fuss decreased over the study period in both groups, the decline by 1 month was greater in the placebo group than probiotic group, with a mean difference in reduction of 46 minutes (95% confidence interval -7 to 99, $P = 0.09$). At 6 months, the groups did not differ for duration of cry or fuss (adjusted mean difference 7 minutes/day, 95% confidence interval -47 to 34, $P = 0.75$).

By 1 month, 40% ($n = 27$) of the treatment group and 48% ($n = 29$) of the placebo group showed at least a 50% reduction in duration of cry or fuss (odds ratio 0.7, 95% confidence interval 0.4 to 1.3, $P = 0.23$). Results from per protocol and intention to treat analyses were similar, as were regression models performed with and without bootstrapping. No treatment effects were evident in the longitudinal analysis using random effects

regression models, and no trends in treatment effect over time were evident.

Secondary outcomes

The two groups were similar on all secondary outcomes at 1 and 6 months (table 3). At 1 month, the probiotic group slept 47 minutes less a day than the placebo group (95% confidence interval -90 to -3, $P=0.04$). Similarly, laboratory analyses of faecal samples at 1 month (treatment group $n=31$, placebo group $n=34$) showed no differences between groups in faecal microbial diversity, calprotectin levels, or *E coli* load. However, infants in both groups with at least a 50% reduction in duration of cry or fuss at 1 month had significantly lower calprotectin levels than non-responders (responders $n=50$, non-responders $n=52$, mean difference 96.6 mg/kg, 95% confidence interval 5.1 to 188.1, $P=0.04$), even though microbial diversity scores and *E coli* load were comparable.

Subgroup analyses

At 1 month, the total duration of cry or fuss was similar between treatment and placebo groups among exclusively breastfed infants (adjusted mean difference 19 minutes, 95% confidence interval -49 to 87, $P=0.57$, $n=50$). Durations of cry or fuss did not differ significantly between the two groups for infants aged more than 6 weeks, and for infants with and without a family history of atopy.

In contrast, formula fed infants in the probiotic group cried or fussed an adjusted mean of 78 minutes more (95% confidence interval 25 to 132, $P=0.005$, $n=71$), and infants aged 6 weeks or less in the probiotic group cried or fussed an adjusted mean of 88 minutes more (16 to 160, $P=0.02$, $n=50$) than corresponding infants in the placebo group. In both cases, this largely resulted from an increase in fussing (adjusted mean difference 63 minutes/day, 21 to 104, $P=0.004$ for formula fed infants, and 67 minutes/day, 8 to 126, $P=0.03$ for infants aged ≤ 6 weeks).

Adherence and adverse events

Reported adherence was 82% (probiotic) and 80% (placebo), with equivalent ingested weights in both groups (mean total product ingested per baby 3.3 g v 3.9 g, $P=0.19$). Analysis of the faecal samples showed that 45% of the probiotic group and none of the placebo group were colonised with *L reuteri* at 1 month post-intervention. No study related adverse events occurred.

Discussion

In this large community cohort of infants with colic sufficiently severe for parents to seek assistance from an emergency care setting, treatment with *Lactobacillus reuteri* did not reduce crying or fussing, nor was it effective in improving infant sleep, maternal mental health, family or infant functioning, or quality of life. In fact, contrary to the hypothesis, infants in the *L reuteri* group fussed more than those in the placebo group at all post-intervention time points except 6 months. This increased fussing occurred only in formula fed infants; *L reuteri* did not affect crying or fussing time in exclusively breastfed infants. *L reuteri* treatment did not lead to changes in infant faecal microbial diversity, *Escherichia coli* colonisation, or calprotectin levels.

Strengths and limitations of this study

This is the largest randomised controlled trial of probiotic intervention in infants with colic to date. It is the first to include formula fed infants and to extend outcomes to infant sleep, maternal mental health, and family functioning. We used rigorous methods and high quality measures; in particular, the well validated Baby Day Diary reduces carer opportunity for recall bias and is sufficiently detailed to permit separated analyses of infant crying versus fussing. Retention and reported adherence rates were high, with the final sample size adequate to achieve the predetermined statistical power and significance.

The trial has some limitations. Most of the infants were recruited from an emergency or urgent care setting; whereas our results may not therefore generalise to infants whose carers do not seek assistance, treatment in this group may be less relevant. Funding permitted completion of laboratory analyses of a subset of faecal samples collected at 1 month only. The low colonisation rate of *L reuteri* in only 45% of the treatment infants may reflect low sensitivity of the assay used. It could also reflect low adherence, although we ascertained good adherence by parental report, and equivalent adherence between the two groups by weights of study bottles before and after ingestion of study product. It remains possible that the infants did not receive a dose of *L reuteri* large enough to establish an effect; however, the trial's probiotic dosing was the same as that of three previous trials reported as effective.³²⁻³⁶ Moreover, the rate of colonisation of *L reuteri* in normal healthy infants given the probiotic is unknown and possibly highly variable. We excluded infants with a likely diagnosis of allergy to cow's milk protein, so our results cannot generalise to this subset of infants.

Interpretation in light of other studies

Despite using the same probiotic strain, formulation, and dosing, our negative trial results contrast with the three previous trials of *L reuteri* in infants with colic,³²⁻³⁶ even when considering only breastfed infants. Our trial also failed to demonstrate positive probiotic effects in formula fed infants with colic. There are several possible reasons for the differences in results. Firstly, our sample size was larger than any previous trial and was adequately blinded, unlike Savino and colleagues' trial.³² Secondly, ours was the only trial that used a validated diary to measure the primary cry or fuss outcome, reducing significantly the likelihood of recall bias and enabling us to uniquely separate out crying and fussing outcomes. Thirdly, the baseline gut microbiota of infants from Melbourne, Australia, could differ from that of Polish and Italian infants, or could be less susceptible to the effects of probiotics. Previous studies have shown that infants with allergic disease have different gut microbiota according to different geographical locations, even among countries with westernised lifestyles.⁴⁴⁻⁴⁸ Fourthly, 60% of our infants had a family history of atopy, in keeping with local epidemiology.⁴⁹ However, the prevalence of a family history of atopy was balanced between groups, and subgroup analysis of infants with and without an atopic family history was similar. Fifthly, it may be that *L reuteri* in formula fed infants is only effective if infants are also receiving a dairy-free diet. Finally, we included infants who were receiving proton pump inhibitors at the start of the trial. However, proton pump inhibitors are ineffective in reducing crying in irritable infants⁵⁰⁻⁵¹ and therefore should have no impact on results.

Unexpectedly, infants in the probiotic group fussed significantly more than infants in the placebo group at all time points from day 7 to 1 month in the adjusted analyses. Further analysis revealed that carers from the placebo group had 40 to 60

minutes/day more contact (carrying/moving) with their infants at all time points, although these differences never reached significance (all $P > 0.05$). The relevance of this is uncertain, as increased contact with carers has been shown to reduce fussing and crying in infants without colic^{52,53} but not in infants with colic.⁵⁴ Previous studies have shown that in exclusively breastfed infants the use of hypoallergenic formula or dairy elimination diets in mothers is somewhat effective in reducing irritability in infants with colic.¹⁰⁻¹² Interestingly, more infants in the placebo group were receiving hypoallergenic formula at 1 month (eight in placebo group v four in probiotic group, $P = 0.08$), and more exclusively breastfed infants in the placebo group had mothers who were on dairy elimination diets at baseline (table 2) and at 1 month (10 in placebo group v 3 in probiotic group, $P = 0.03$). However, these numbers were all small, and in further analyses the results were not altered by controlling for dairy exclusion and use of hypoallergenic formulas.

In our preliminary analysis of faecal samples, calprotectin levels, although unrelated to trial arm, were lower in those with resolved colic. This finding is in keeping with a previous small study showing faecal calprotectin levels to be higher in infants with colic than in those without,⁸ but contrary to a larger study showing no differences in faecal calprotectin levels between infants with and without colic.⁹ Considering that faecal calprotectin levels are associated with intestinal neutrophilic inflammation, it is conceivable that the more than 50% reduction in crying or fussing may reflect lower levels of intestinal inflammation by a mechanism unrelated to probiotic ingestion. These can be further explored by better designed studies investigating faecal calprotectin levels and infant crying.

When the results of our trial are added to the meta-analysis of previous randomised controlled trials of *L reuteri* for the management of infant colic,³¹ the overall effectiveness of the probiotic is diminished: mean difference 48 minutes/day (95% confidence interval -84.8 to -11.3) compared with 68 minutes/day (-99.8 to -35.6); figures 3↓ and 4↓). However, it should be noted that the meta-analysis puts significant weighting on the study by Savino and colleagues,³² which was an unblinded, open label trial.

Conclusions and future directions

Based on the null findings from this large, rigorous trial, and the results from the updated meta-analysis, which has potential biases, we recommend caution in formulating recommendations regarding use of *L reuteri* DSM 17938 for reducing crying or fussing in breastfed, and particularly in formula fed, infants with colic. Currently, there are at least five other similar randomised controlled trials taking place in different parts of the world.⁵⁵ It will be valuable to pool data from all existing and ongoing rigorous trials into an individual patient level meta-analysis to investigate with greater certainty which subgroups of infants with colic, if any, could benefit from probiotics.

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Contributors: All authors had full access to all of the data (including statistical reports and tables) in the study and can take responsibility for the integrity of the data and the accuracy of the data analysis. VS is the guarantor, had final responsibility for the decision to submit for publication, and takes overall responsibility for all aspects of the trial and this manuscript. VS, HH, and MW conceived the trial with MT, RH, and FM. VS, HH, and MW designed the intervention, with advice from MT, RH, and RGB, who also advised on measures and their interpretation. FM advised on statistical issues. AS contributed to recruitment procedures. MT advised on laboratory outcome measures and their interpretation, with input from CS and MN. MN developed and performed laboratory assays with advice and oversight from CS. All authors contributed to, read, and approved the final manuscript.

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Ethical approval: This trial was approved by the Royal Children's Hospital human research ethics committee (HREC 30111). All participants gave written informed consent before taking part.

Data sharing: The study's technical appendix is available with open access at www.rch.org.au/cchc/for_researchers/Baby_Biotics/. The statistical code is available, and anonymous patient level data may be available, from the corresponding author (valerie.sung@rch.org.au) subject to the study's material transfer agreement. Consent from participants for data sharing was not obtained but the presented data are anonymised and risk of identification is low.

Transparency: VS affirms that the manuscript is an honest, accurate, and transparent account of the study being reported; that no important aspects of the study have been omitted; and that any discrepancies from the study as planned (and, if relevant, registered) have been explained.

- 1 Wake M, Morton-Allen E, Poulakis Z, Hiscock H, Gallagher S, Oberklaid F. Prevalence, stability, and outcomes of cry-fuss and sleep problems in the first 2 years of life: prospective community-based study. *Pediatrics* 2006;117:836-42.
- 2 Wessel MA, Cobb JC, Jackson EB, Harris GS, Detwiler AC. Paroxysmal fussing in infancy, sometimes called "colic." *Pediatrics* 1954;14:421-34.
- 3 McMahon C, Barnett B, Kowalenko N, Tennant C, Don N. Postnatal depression, anxiety and unsettled infant behaviour. *Aust N Z J Psychiatry* 2001;35:581-8.

What is already known on this topic

Previous small trials suggest that the probiotic *Lactobacillus reuteri* effectively treats colic in breastfed infants
 These studies, however, had limitations as they examined a highly selective group of infants with colic
 The effects of *L reuteri* on formula fed infants with colic are unknown

What this study adds

L reuteri treatment did not reduce crying or fussing in infants with colic, nor was it effective in improving infant sleep, maternal mental health, family or infant functioning, or quality of life
 Probiotics therefore cannot be routinely recommended for all infants with colic
 Further research is needed to identify which subgroups of infants with colic may benefit from probiotics

- 4 Howard CR, Lanphear N, Lanphear BP, Eberly S, Lawrence RA. Parental responses to infant crying and colic: the effect on breastfeeding duration. *Breastfeed Med* 2006;1:146-55.
- 5 Barr RG. Preventing abusive head trauma resulting from a failure of normal interaction between infants and their caregivers. *Proc Natl Acad Sci USA* 2012;109(Suppl 2):17294-301.
- 6 Morris S, James-Roberts IS, Sleep J, Gillham P. Economic evaluation of strategies for managing crying and sleeping problems. *Arch Dis Childhood* 2001;84:15-9.
- 7 Barr RG. Excessive crying. In: Sameroff AJ, Lewis M, Miller SM, eds. Handbook of developmental psychopathology. Kluwer Academic/Plenum Press, 2000:327-50.
- 8 Rhoads JM, Fatheree NY, Norori J, Liu Y, Lucke JF, Tyson JE, et al. Altered fecal microflora and increased fecal calprotectin in infants with colic. *J Pediatr* 2009;155:823-8.e1.
- 9 Olafsdottir E, Aksnes L, Fluge G, Berstad A. Faecal calprotectin levels in infants with infantile colic, healthy infants, children with inflammatory bowel disease, children with recurrent abdominal pain and healthy children. *Acta Paediatr* 2002;91:45-50.
- 10 Cohen-Silver J, Ratnapalan S. Management of infantile colic: a review. *Clin Pediatr* 2009;48:14-7.
- 11 Garrison MM, Christakis DA. A systematic review of treatments for infant colic.[see comment]. *Pediatrics* 2000;106:184-90.
- 12 Lucassen PL, Assendelft WJ. Systematic review of treatments for infant colic. *Pediatrics* 2001;108:1047-8.
- 13 National Institute for Health and Care Excellence guidelines. Colic—infantile. Clinical knowledge summaries. 2014. <http://cks.nice.org.uk/colic-infantile#management>.
- 14 Heine RG. Cow's milk allergy and lactose malabsorption in infants with colic. *J Pediatr Gastroenterol Nutr* 2013;57(Suppl 1):25-7.
- 15 Ghosh S, Barr RG. Colic and gas. In: Walker W, Goulet O, Kleinman R, Sherman P, Schneider B, Sanderson I, eds. Pediatric gastrointestinal disease. BC Decker, 2004:210-24.
- 16 Weissbluth M, Christoffel KK, Davis AT. Treatment of infantile colic with dicyclomine hydrochloride. *J Pediatr* 1984;104:951-5.
- 17 Illingworth RS. Evening colic in infants: a double-blind trial of dicyclomine hydrochloride. *Lancet* 1959;2:1119-20.
- 18 Grunseit F. Evaluation of the efficacy of dicyclomine hydrochloride ('Merbentyl') syrup in the treatment of infant colic. *Curr Med Res Opin* 1977;5:258-61.
- 19 Williams J, Watkins-Jones R. Dicyclomine: worrying symptoms associated with its use in some small babies. *BMJ* 1984;288:901.
- 20 De Weerth C, Fuentes S, Puylaert P, de Vos WM. Intestinal microbiota of infants with colic: development and specific signatures. *Pediatrics* 2013;131:e550-8.
- 21 Savino F, Cordisco L, Tarasco V, Calabrese R, Palumeri E, Matteuzzi D. Molecular identification of coliform bacteria from colicky breastfed infants. *Acta Paediatr* 2009;98:1582-8.
- 22 Savino F, Cresi F, Pautasso S, Palumeri E, Tullio V, Roana J, et al. Intestinal microflora in breastfed colicky and non-colicky infants. *Acta Paediatr* 2004;93:825-9.
- 23 Partty A, Kalliomaki M, Endo A, Salminen S, Isolauri E. Compositional development of Bifidobacterium and Lactobacillus microbiota is linked with crying and fussing in early infancy. *PLoS One* 2012;7.
- 24 Penna FJ, Peret LA, Vieira LQ, Nicoli JR. Probiotics and mucosal barrier in children. *Curr Opin Clin Nutr Metab Care* 2008;11:640-4.
- 25 Grzeskowiak L, Gronlund MM, Beckmann C, Salminen S, von Berg A, Isolauri E. The impact of perinatal probiotic intervention on gut microbiota: double-blind placebo-controlled trials in Finland and Germany. *Anaerobe* 2012;18:7-13.
- 26 Duchmann R. The role of probiotics and antibiotics in regulating mucosal inflammation. *Adv Exp Med Biol* 2006;579:219-26.
- 27 Lutgendorff F, Akkermans LM, Soderholm JD. The role of microbiota and probiotics in stress-induced gastro-intestinal damage. *Curr Mol Med* 2008;8:282-98.
- 28 Mohan R, Koebnick C, Schildt J, Schmidt S, Mueller M, Pössner M, et al. Effects of Bifidobacterium lactis Bb12 supplementation on intestinal microbiota of preterm infants: a double-blind, placebo-controlled, randomized study. *J Clin Microbiol* 2006;44:4025-31.
- 29 Mentula S, Tuure T, Koskenala R, Korpela R, Kononen E. Microbial composition and fecal fermentation end products from colicky infants—a probiotic supplementation pilot. *Microb Ecol Health Dis* 2008;20:37-47.
- 30 Savino F, Cordisco L, Tarasco V, Locatelli E, Di Gioia D, Oggero R, et al. Antagonistic effect of Lactobacillus strains against gas-producing coliforms isolated from colicky infants. *BMC Microbiol* 2011;11:157.
- 31 Sung V, Collett S, De Gooyer T, Hiscoc H, Tang ML, Wake M. Probiotics to prevent or treat excessive infant crying: Systematic review and meta-analysis. *JAMA Pediatrics* 2013;167:1150-7.
- 32 Savino F, Pelle E, Palumeri E, Oggero R, Miniero R. Lactobacillus reuteri (American type culture collection strain 55730) versus simethicone in the treatment of infantile colic: a prospective randomized study. *Pediatrics* 2007;119:e124-30.
- 33 Savino F, Cordisco L, Tarasco V, Palumeri E, Calabrese R, Oggero R, et al. Lactobacillus reuteri DSM 17 938 in infantile colic: a randomized, double-blind, placebo-controlled trial. *Pediatrics* 2010;126:e526-33.
- 34 Sung V, Hiscoc H, Tang ML, Mensah FK, Heine RG, Stock A, et al. Probiotics to improve outcomes of colic in the community: protocol for the Baby Biotics randomised controlled trial. *BMC Pediatr* 2012;12:135.
- 35 Barr RG, Kramer MS, Leduc DG, Boisjoly C, McVey-White L, Pless IB. Parental diary of infant cry and fuss behavior. *Arch Dis Child* 1988;63:380-7.
- 36 Szajewska H, Gyrzduk E, Horvath A. Lactobacillus reuteri DSM 17938 for the management of infantile colic in breastfed infants: a randomized, double-blind, placebo-controlled trial. *J Pediatr* 2013;162:257-62.
- 37 Cox JL, Holden JM, Sagovski R. Detection of postnatal depression: development of a 10-item Edinburgh postnatal depression scale. *Br J Psychiatry* 1987;150:1172-5.
- 38 Varni JW, Limbers CA, Nieghors K, Schulz K, Lieu JEC, Heffer RW, et al. The PedsQL infant scales: feasibility, internal consistency, reliability, and validity in healthy and ill infants. *Qual Life Res* 2010; published online 22 Aug. doi: 10.1007/s11136-010-9730-5.
- 39 Varni JW, Sherman SA, Burwinkle TM, Dickinson PE, Dixon P. The PedsQL™ family impact module: preliminary reliability and validity. *Health Qual Life Outcomes* 2004;2:55.
- 40 Hawthorne G, Richardson J, Atherton Day M. A comparison of the Assessment of Quality of Life (AqoL) with four other generic utility instruments. *Ann Med* 2001;33:358-70.
- 41 Osborne CA, Rees GN, Bernstein Y, Janssen PH. New threshold and confidence estimates for terminal restriction fragment length polymorphism analysis of complex bacterial communities. *Appl Environ Microbiol* 2006;72:1270-8.
- 42 Chern EC, Siefiring S, Paar J, Doolittle M, Haugland RA. Comparison of quantitative PCR assays for Escherichia coli targeting ribosomal RNA and single copy genes. *Letts Appl Microbiol* 2011;52:298-306.
- 43 CALPRO. CalproLab ELISA kit. 2012. www.calpro.no/uploads/pdf/110831_CalproLab%20ALP%20ELISA.pdf.
- 44 Ouwehand AC, Isolauri E, He F, Hashimoto H, Benno Y, Salminen S. Differences in Bifidobacterium flora composition in allergic and healthy infants. *J Allergy Clin Immunol* 2001;108:144-5.
- 45 Stsepetova J, Sepp E, Julge K, Vaughan E, Mikelsaar M, de Vos WM. Molecularly assessed shifts of Bifidobacterium ssp and less diverse microbial communities are characteristic of 5-year-old allergic children. *FEMS Immunol Med Microbiol* 2007;51:260-9.
- 46 Songjinda P, Nakayama J, Tateyama A, Tanaka S, Tsubouchi M, Kiyohara C, et al. Differences in developing intestinal microbiota between allergic and non-allergic infants: a pilot study in Japan. *Biosci Biotechnol Biochem* 2007;71:2338-42.
- 47 Suzuki S, Shimojo N, Tajiri Y, Kumemura M, Kohno Y. Differences in the composition of intestinal Bifidobacterium species and the development of allergic diseases in infants in rural Japan. *Clin Exp Allergy* 2007;37:506-11.
- 48 Gore C, Munro K, Lay C, Bibiloni R, Morris J, Woodcock A, et al. Bifidobacterium pseudocatenulatum is associated with atopic eczema: a nested case-control study investigating the fecal microbiota of infants. *J Allergy Clin Immunol* 2008;121:135-40.
- 49 Osborne NJ, Koplin JJ, Martin PE, Gurrin LC, Thiele L, Tang ML, et al. The HealthNuts population-based study of paediatric food allergy: validity, safety and acceptability. *Clin Exp Allergy* 2010;40:1516-22.
- 50 Moore DJ, Tao BS, Lines DR, Hirte C, Heddle ML, Davidson GP. Double-blind placebo-controlled trial of omeprazole in irritable infants with gastroesophageal reflux. *J Pediatr* 2003;143:219-23.
- 51 Jordan B, Heine RG, Meehan M, Catto-Smith AG, Lubitz L. Effect of antireflux medication, placebo and infant mental health intervention on persistent crying: a randomized clinical trial. *J Paediatr Child Health* 2006;42:49-58.
- 52 St James-Roberts I, Alvarez M, Csipke E, Abramsky T, Goodwin J, Sorgenfrei E. Infant crying and sleeping in London, Copenhagen and when parents adopt a "proximal" form of care. *Pediatrics* 2006;117:e1146-55.
- 53 St James-Roberts I, Hurry J, Bowyer J, Barr RG. Supplementary carrying compared with advice to increase responsive parenting as interventions to prevent persistent infant crying. *Pediatrics* 1995;95:381-8.
- 54 Barr RG, McMullan SJ, Spiess H, Leduc DG, Yaremko J, Barfield R, et al. Carrying as colic "therapy": a randomized controlled trial. *Pediatrics* 1991;87:623-30.
- 55 MetaregisterOfControlledTrials. Registered trials numbers ISRCTN95287767, NCT01067027, NCT01279265, NCT01541046, NCT01017991. 2012. www.controlled-trials.com/mrct/search.html.

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Tables

Table 1 | Outcome measures (adapted from Sung et al³⁴)

Outcomes	Days					Month	Measure	Additional information
	0	7	14	21	28	6		
Primary outcome:								
Daily cry/fuss time (min/day)	—	—	—	—	Y	—	Baby's Day Diary	Validated measure of infant crying, fussing, sleep, and feeding. Records these behaviours in 5 minute epochs over 24 hours. ³⁵ At baseline, parents record in study diary for 24 hours. At other time points, diary filled in over 48 hours (to reduce daily variability in infant behaviour while taking into account potential burden to families from filling in diary for prolonged periods)
Secondary outcomes:								
Daily cry/fuss time (min/day)	Y	Y	Y	Y	—	Y	Baby's Day Diary	
Daily cry/fuss episodes	Y	Y	Y	Y	Y	Y	Baby's Day Diary	
Infant sleep duration (min/day)	Y	Y	Y	Y	Y	Y	Baby's Day Diary	
Maternal mental health scores	Y	—	—	—	Y	Y	Edinburgh postnatal depression subscale	10 item validated questionnaire to screen for depression in postpartum period, with higher scores indicating poorer mental health. ³⁷ Scores validated to detect postnatal depression in community (≥ 10) and clinical (≥ 12) settings
Infant functioning scores	—	—	—	—	—	Y	Paediatric quality of life inventory infant subscale	36 item validated questionnaire to measure infant physical, emotional, social, and cognitive functioning ³⁸
Family functioning scores	Y	—	—	—	Y	Y	Paediatric quality of life inventory family impact subscale	5 item validated questionnaire to assess family functioning, with higher scores indicating better family functioning ³⁹
Parent quality adjusted life years scores	Y	—	—	—	Y	Y	Assessment of quality of life (AQoL-4R)	12 item validated questionnaire to assess health economic parent quality of life ⁴⁰
Infant faecal microbial diversity*	Y	—	—	—	Y	—	16SrDNA amplification (T-RFLP)	Terminal restriction fragment length polymorphism, a molecular method to investigate diversity within bacterial communities, given as diversity score, with higher scores indicating more microbial diversity ⁴¹
Infant faecal <i>Escherichia coli</i> colonisation(cfu/mL)*	Y	—	—	—	Y	—	Quantitative polymerase chain reaction	Molecular method to detect and measure presence of a marker gene of <i>E coli</i> ⁴²
Infant faecal calprotectin (mg/kg)†	Y	—	—	—	Y	—	Enzyme linked immunosorbent assay	To detect presence of calprotectin, a marker of gut inflammation ⁴³

cfu=colony forming units.

*Funding allowed analysis of a subset of faecal samples collected at one month post-intervention only.

†Funding allowed analysis of first 102 faecal samples collected at one month post-intervention only.

Table 2| Baseline characteristics. Values are numbers (percentages) unless stated otherwise

Characteristics	No	Probiotic group	No	Placebo group
Boys	85	37 (44)	82	48 (59)
Mean (SD) age at study entry (weeks)		7.5 (2.9)		6.9 (2.5)
Mean (SD) birth weight (g)		3272 (406)		3426 (421)
Mean (SD) gestation (weeks)		38.7 (1.5)		39.1 (1.4)
Caesarean delivery		35 (41)		32 (39)
Family history of atopy		51 (60)		50 (61)
Exclusive breast feeding at entry:		33 (39)		35 (43)
Dairy exclusion	33	7 (22)	35	12 (34)
Formula use at entry:	85	52 (61)	82	47 (57)
Hypoallergenic formula use	52	7 (13)	47	8 (17)
Probiotic containing formula use	52	18 (35)	47	15 (32)
Any breast feeding at entry:	85	55 (65)	82	54 (66)
Probiotic use in any breastfeeding mother	55	12 (22)	54	10 (19)
Supplementary probiotic use in infants at entry	85	6 (7)	82	3 (4)
Proton pump inhibitor use in infants at entry	80	21 (26)	68	24 (35)
Mean (SD) total daily cry/fuss time (min/day):	75	328 (152)	65	329 (126)
Fuss time		153 (107)		163 (94)
Cry time		175 (115)		166 (93)
Mean (SD) daily cry/fuss episodes		10.3 (4.9)		11.8 (6.3)
Mean (SD) infant sleep duration (min/day)		728 (161)		745 (141)
Mean (SD) maternal mental health (EPDS score)		10.5 (4.7)		10.6 (4.6)
Mean (SD) family functioning (PedsQL score)		71.2 (23.9)		75.2 (21.3)
Mean (SD) parent quality adjusted life years (AQoL score)		0.74 (0.2)		0.74 (0.2)

EPDS=Edinburgh postnatal depression scale; PedsQL=paediatric quality of life inventory; AQoL=assessment of quality of life.

Table 3| Outcomes at 1 and 6 months in treatment and placebo groups

Outcomes	Mean (SD)				Adjusted mean difference* (95% CI)	P value	Median (interquartile range)		P value†
	No	Probiotic	No	Placebo			Probiotic	Placebo	
1 month:									
Total daily cry or fuss time (min/day)	67	229 (137)	60	191 (103)	49 (8 to 90)	0.02	203 (130-295)	166 (128-265)	0.14
Fuss time (min/day)	67	153 (109)	60	112 (78)	52 (19 to 84)	0.002	125 (75-210)	106 (51-150)	0.04
Cry time (min/day)	67	76 (82)	60	79 (77)	-2 (-28 to 24)	0.86	56 (13-105)	63 (30-100)	0.4
Daily cry or fuss episodes	67	7.5 (4.7)	59	8.1 (4.9)	0.4 (-1.1 to 1.8)	0.62	7 (4-11)	7 (5-12)	0.52
Infant sleep duration (min/day)	67	800 (141)	60	842 (119)	-47 (-90 to -3)	0.04	818 (718-903)	857 (772-921)	0.08
Maternal mental health (EPDS score)	79	8.2 (4.7)	75	7.9 (4.9)	0.6 (-0.7 to 1.9)	0.36	8 (4-12)	8 (4-11)	0.63
Family functioning (PedsQL score)	79	69.8 (21.6)	75	71.1 (20.8)	-0.7 (-6.1 to 4.7)	0.80	70 (56-85)	75 (60-85)	0.70
Parent quality adjusted life years (AQoL score)	79	0.7 (0.2)	74	0.8 (0.2)	0.0 (-0.1 to 0.0)	0.11	0.7 (0.6-0.9)	0.8 (0.7-0.9)	0.19
Infant faecal microbial diversity score	28	32.8 (1.4)	27	30.4 (1.0)	-2.5‡ (-5.9 to 1.0)	0.16	31 (28-37)	31 (27-35)	0.34
Infant faecal <i>Escherichia coli</i> colonisation (cfu/mL)	31	3.1 (5.3)×10 ⁷	34	2.6 (5.1)×10 ⁷	-0.5‡ (-3.1 to 2.1×10 ⁷)	0.70	0.8 (0.9-3.0)×10 ⁷	0.5 (0.0-2.4)×10 ⁷	0.31
Infant faecal calprotectin (mg/kg)	53	230.0 (36.5)	49	197.4 (28.9)	-32.6‡ (-125.9 to 60.7)	0.49	135 (74-259)	114 (73-242)	0.63
6 months:									
Total daily cry or fuss time (min/day)	65	122 (118)	58	131 (111)	-7 (-47 to 34)	0.75	85 (55-140)	105 (60-160)	0.27
Fuss time (min/day)	65	84 (77)	58	84 (83)	0 (-29 to 29)	0.99	63 (30-100)	61 (35-105)	0.94
Cry time (min/day)	65	38 (56)	58	47 (52)	-7 (-27 to 13)	0.49	18 (0-45)	29 (10-75)	0.13
Daily cry or fuss episodes	65	5.8 (5.0)	58	6.6 (4.4)	-0.1 (-1.8 to 1.5)	0.87	4 (3-8)	6 (3-9)	0.08
Infant sleep duration (min/day)	65	776 (185)	58	783 (167)	1 (-66 to 68)	0.97	810 (720-880)	807 (730-870)	0.98
Maternal mental health (EPDS score)	69	6.5 (4.5)	63	7.1 (4.7)	-0.1 (-1.6 to 1.3)	0.84	6 (3-9)	6 (3-11)	0.44
Family functioning (PedsQL score)	69	78.4 (19.4)	63	76.0 (21.7)	1.0 (-5.5 to 7.6)	0.76	80 (70-95)	80 (60-100)	0.70
Infant functioning (PedsQL score)	69	77.7 (11.4)	63	76.3 (11.1)	0.9§ (-3.1 to 4.8)	0.66	78 (72-84)	75 (69-84)	0.32
Parent quality adjusted life years (AQoL score)	69	0.8 (0.2)	63	0.8 (0.2)	-0.1 (-0.1 to 0.0)	0.07	0.8 (0.7-0.9)	0.8 (0.7-1.0)	0.22

cfu=colony forming units; EPDS=Edinburgh postnatal depression scale; PedsQL=paediatric quality of life inventory; AQoL=assessment of quality of life.

*Adjusted for sex, age, delivery mode, feeding type, family history of atopy, and respective baseline variable.

†Two sample Wilcoxon rank sum test.

‡Unadjusted analyses of available faecal samples.

§Adjusted for sex, age, delivery mode, feeding type, and family history of atopy.

Figures

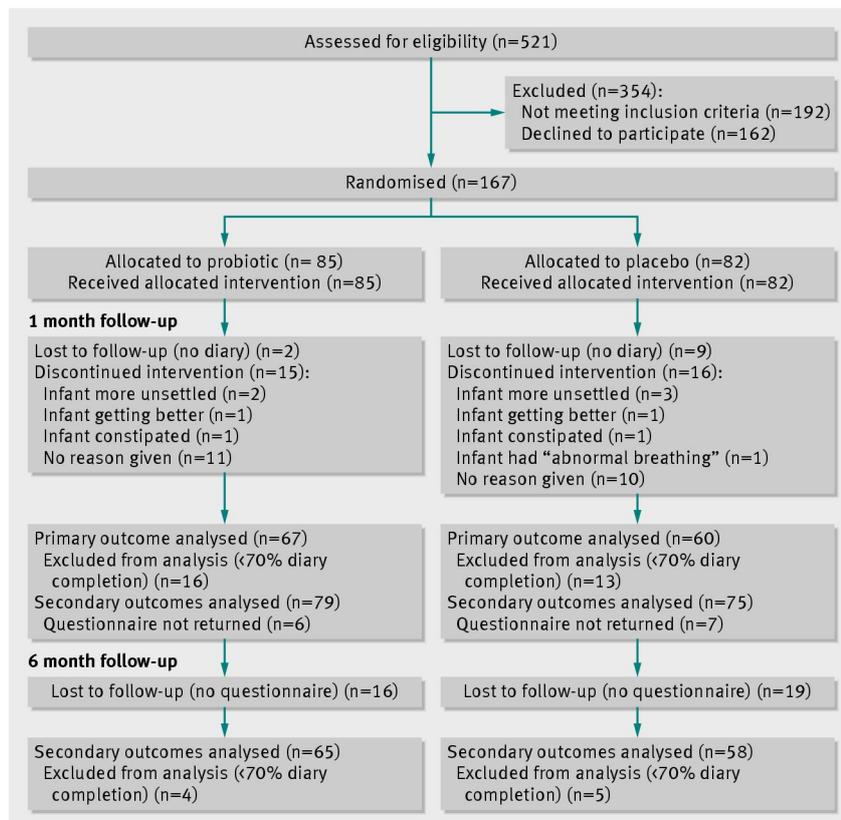


Fig 1 Participant flow through study

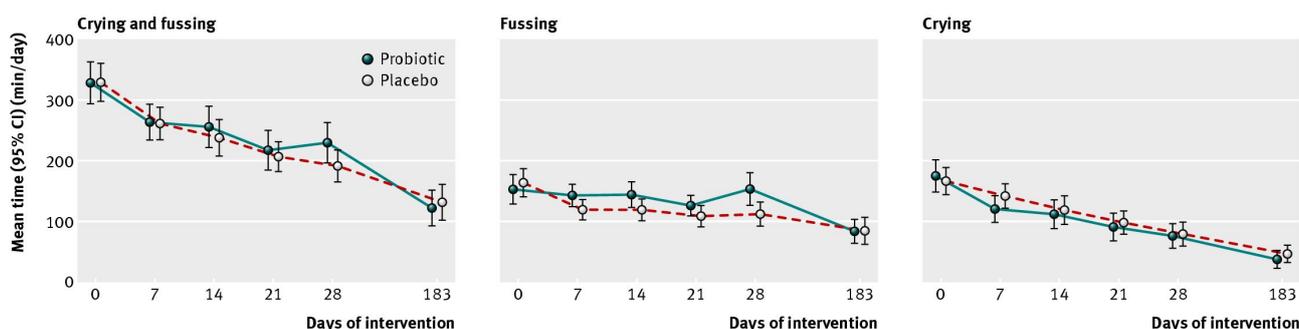


Fig 2 Daily duration of cry or fuss over study period and at 6 month follow-up. Day 28=1 month; day 183=6 months

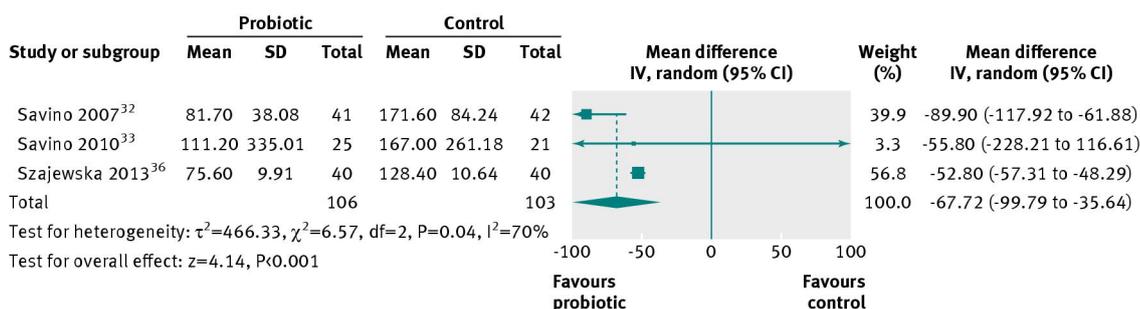


Fig 3 Meta-analysis of previous randomised controlled trials of probiotics for management of infant colic (outcomes at 21 days post-intervention)³¹

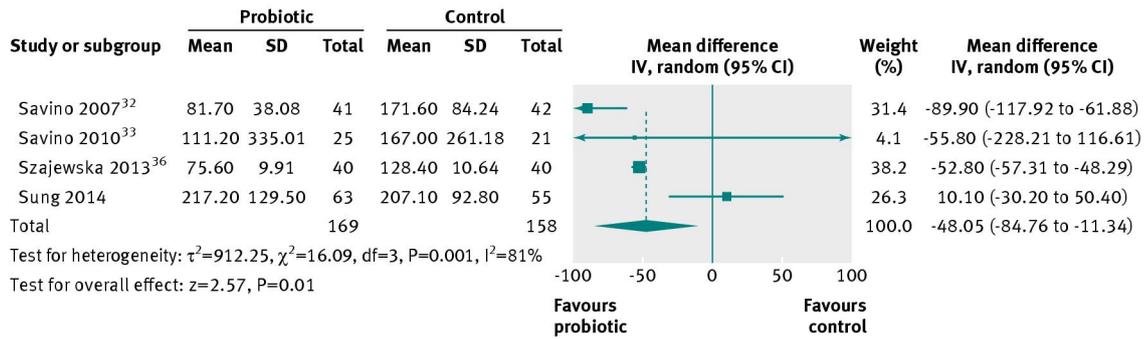


Fig 4 Meta-analysis of previous randomised controlled trials of probiotics for management of infant colic with addition of results from this study